DOI: 10.1002/agg2.20378

#### ORIGINAL ARTICLE

Agrosystems

# Late postemergence glufosinate-based programs for glyphosate-resistant Palmer amaranth control in dicamba/glufosinate/glyphosate-resistant soybean

Vipan Kumar<sup>1</sup> 💿 🕴 Jatinder S. Aulakh<sup>2</sup> 🕴 Rui Liu<sup>3</sup> 👘 Amit J. Jhala<sup>4</sup>

<sup>1</sup>Soil and Crop Sciences Section, School of Integrative Plant Science, Cornell University, Ithaca, New York, USA

<sup>2</sup>Connecticut Agricultural Experiment Station, Windsor, Connecticut, USA

<sup>3</sup>Irrigated Agriculture Research and Extension Center, Washington State University, Prosser, Washington, USA

<sup>4</sup>Department of Agronomy and Horticulture, University of Nebraska, Lincoln, Nebraska, USA

#### Correspondence

Vipan Kumar, Soil and Crop Sciences Section, School of Integrative Plant Science, Cornell University, 1115 Bradfield Hall, Ithaca, NY 14853, USA. Email: vk364@cornell.edu

Assigned to Associate Editor Kurt Matthew Vollmer.

#### Abstract

Glyphosate-resistant (GR) Palmer amaranth (Amaranthus palmeri S. Watson) is widespread in the Central Great Plains. Introduction of newly developed dicamba/glufosinate/glyphosate (DGG)-resistant soybean varieties allows postemergence (POST) applications of dicamba and glufosinate for in-season control of GR Palmer amaranth. Limited information exists on the effectiveness of glufosinate applied late-POST for tall (70-90 cm) GR Palmer amaranth control in DGG-resistant soybean. The objectives of this study were to (1) determine the effectiveness of late-POST glufosinate-based programs for GR Palmer amaranth control, and (2) determine the impact of those programs on soybeans grain yields. Ten glufosinatebased programs were tested in a field study at Kansas State University Agricultural Research Center near Hays, Kansas. Results indicated that single (655 or 737 g ha<sup>-1</sup>) and all sequential (594 followed by [fb] 594, 655 fb 594, and 737 fb 594 g ha<sup>-1</sup>) applications (7-days apart) of glufosinate provided 87%-93% control of GR Palmer amaranth 28 days after last POST (DALPOST). Palmer amaranth control with single late-POST application of glufosinate (594 g  $ha^{-1}$ ) or glufosinate plus S-metolachlor did not exceed 84% at 28 DALPOST. Majority of the evaluated programs reduced shoot dry weights of GR Palmer amaranth by 83%-91%. The least control (11%) and shoot dry weight reduction (33%) of GR Palmer amaranth were observed with glyphosate fb glyphosate. Glufosinate-based programs resulted in soybean grain yield of 626–701 kg ha<sup>-1</sup>. These results conclude that glufosinate applied late-POST may provide effective control of tall GR Palmer amaranth in DGG-resistant soybeans.

#### **1** | INTRODUCTION

Abbreviations: DALPOST, days after last POST; DGG, dicamba/glufosinate/glyphosate; EPOST, early POST; fb, followed by; GR, glyphosate-resistant; POST, postemergence; PRE, preemergence. Palmer amaranth (*Amaranthus palmeri* L. Watts.) is among the most troublesome summer annual broadleaf weeds in agronomic crops in the United States (Van Wychen, 2017), including Kansas (Kumar et al., 2020). Palmer amaranth is a member of the pigweed (Amaranthaceae) family and is native

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made. © 2023 The Authors. *Agrosystems, Geosciences & Environment* published by Wiley Periodicals LLC on behalf of Crop Science Society of America and American Society of Agronomy.

to the southwest United States and northwest Mexico (Sauer, 1957; Ward et al., 2013). Palmer amaranth possesses several unique characteristics, including an extended emergence period (early May to late September), rapid plant growth rate, dioecy (male and female flowers are on separate plants), high outcrossing potential (can outcross within and between species in the pigweed family) and high genetic diversity within and among populations, and prolific seed production (up to 0.6 million seeds per female plant) (Adhikary & Pratt, 2015; Horak & Loughin, 2000; Keeley et al., 1987; Steckel et al., 2004; Ward et al., 2013). Season-long competition from Palmer amaranth at a density of 10 plants m<sup>-2</sup> reduced soybean grain yield by 68% in Arkansas, USA (Klingaman & Oliver, 1994).

Due to greater genetic diversity and prolific seed production, Palmer amaranth exhibits a high propensity to evolve herbicide resistance (Heap, 2023; Ward et al., 2013). For instance, Palmer amaranth populations resistant to herbicides that inhibit acetolactate synthase (ALS), 5-enolpyruvyl shikimate-3-phosphate synthase (EPSPS), microtubule assembly (MTA), photosystem II (PS II), 4-hydroxyphenyl pyruvate dioxygenase (HPPD), and protoporphyrinogen oxidase (PPO) have been widely reported (Heap, 2023). Glyphosate-resistant (GR) Palmer amaranth was first confirmed in Kansas in 2011 (Heap, 2023); as of 2023, it is widely spread in several counties in Kansas (Kumar et al., 2020). More recently, Palmer amaranth with resistance to 2,4-D in Kansas, to dicamba in Tennessee, and to S-metolachlor and glufosinate in Arkansas have also been reported (Brabham et al., 2019; Foster & Steckel, 2022; Kumar et al., 2019; Priess et al., 2022). In addition, multiple resistance to five to six herbicide sites of action has also been reported in Palmer amaranth populations in Kansas and Arkansas (Heap, 2023; Kumar et al., 2019, 2020).

Effective control of GR Palmer amaranth in no-till dryland soybean primarily relies on two-pass herbicide programs (residual herbicides applied at planting followed by an early POST [EPOST] herbicide) (Kumar et al., 2020). However, escapes from EPOST and the late-season emergence pattern of Palmer amaranth are generally observed in the Central Great Plains region (Liu et al., 2022). Few escaped or late-emerged female Palmer amaranth plants can produce a significant amount (>50,000 seeds per plant) of seeds (Kumar, Liu, Jhala, et al., 2021) and can substantially contribute to the soil seed bank (Jha & Norsworthy, 2009). Therefore, management of those late-emerged Palmer amaranth cohorts with mid- to late-season herbicide applications is crucial in soybean fields where no preemergence (PRE) herbicide was applied, and postemergence (POST) herbicide is the only option in a no-till dryland production system (de Sanctis et al., 2021).

Dicamba/glufosinate/glyphosate (DGG)-resistant soybean (XtendFlex, Bayer CropScience) was commercially launched

#### **Core Ideas**

- Late-POST glufosinate-based herbicide programs were evaluated in dicamba/glufosinate/glyphosate (DGG)-resistant soybeans.
- Single (655 or 737 g ha<sup>-1</sup>) and all sequential late-POST glufosinate programs provided 87%–93% control of GR Palmer amaranth.
- The majority of glufosinate-based programs reduced shoot dry weights of GR Palmer amaranth by 83%–91%.
- Late-POST glufosinate programs had 56%–61% higher soybean yield compared to glyphosate followed by glyphosate.

in 2021 in the United States and Canada. The adoption of DGG-resistant soybean allows growers to use single or sequential applications of low-volatile dicamba formulations and glufosinate for in-season control of GR weeds such as waterhemp (Amaranthus tuberculatus L.) and Palmer amaranth (Jones et al., 2022; Meyer et al., 2015). However, the cut-off dates for POST dicamba applications on DGG-resistant soybean vary from state to state according to recent changes approved by the EPA. For the 2023 growing season, no POST dicamba applications are allowed on DGG-resistant soybean after June 12 or V4 growth stage (whichever comes first) in Iowa, Illinois, Indiana, and south of Interstate 94 in Minnesota (Jenkins, 2023). Furthermore, the cut-off date for POST dicamba applications on DGG-resistant soybeans is June 20 for South Dakota and June 30 for all other states (Jenkins, 2023). These restrictions leave glufosinate as a sole POST herbicide option for late-season control of GR weeds in DGG-resistant soybean.

Glufosinate is a contact, broad-spectrum POST herbicide that inhibits the glutamine synthetase enzyme in plants, which eventually lead to cell membrane disruption and necrosis (Haas & Muller, 1987; Hinchee et al., 1993). Glufosinate can be applied in single or sequential applications from emergence up to R1 growth stage of DGG-resistant soybean and its efficacy depends on the growth stage of the weed (Anonymous, 2019; Aulakh & Jhala, 2015). However, soybean growers often apply POST herbicides when Palmer amaranth is in variable heights (de Sanctis et al., 2021). Furthermore, temperature and relative humidity are known to influence the efficacy of glufosinate (Anderson et al., 1993). The objectives of this research were to (1) determine the efficacy of glufosinate-based herbicide programs applied late-POST on GR Palmer amaranth (70-90 cm tall) control, and (2) the impact of those programs on GR Palmer amaranth biomass

and DGG-resistant soybean grain yields in no-till dryland Central Grain Plains region.

#### **MATERIALS AND METHODS** 2

#### 2.1 Field study

A field study was conducted at Kansas State University Agricultural Research Center (KSU-ARC) near Hays, Kansas (38.86177°N, 99.33396°W) in 2022 growing season. Soil type at the study site was Roxbury silt loam with pH of 7.8 and organic matter of 2.0%. The study site was under notill dryland system with a typical 3-year crop rotation wheat (Triticum aestivum L.) followed by (fb) summer crop fb fallow for >10 years. Paraquat at 560 g ha<sup>-1</sup> was applied at the study site for control of existing weeds such as kochia (Bassia scoparia L.), blue mustard (Chorispora tenella L.), and horseweed (Conyza canadensis L.) before soybean planting. The experimental site had a natural seed bank of GR Palmer amaranth.

Study was conducted in a randomized complete block design (RCBD) with four replications. A DGG-resistant soybean (XtendFlex<sup>â</sup> soybean) variety 'AG37XF1' was planted at 387,543 seeds  $ha^{-1}$  in 76-cm spaced rows (4 rows plot<sup>-1</sup>) in  $3.0 \times 9.1$  m plots on May 25, 2022. A blanket treatment of glyphosate (1260 g ae  $ha^{-1}$ ) along with ammonium sulfate (AMS) at 2% wt/v was applied at V2-V3 growth stage of soybean to control green foxtail. All emerged Palmer amaranth plants had shown little to no injury to glyphosate, further indicating the presence of GR Palmer amaranth population at the study site (data not shown). All late-POST programs (Table 1) of glufosinate were applied with a  $CO_2$ -operated backpack sprayer equipped with flat-fan nozzles (Turbo Teejet XR 110015; Spraving Systems Co.), calibrated to deliver 140 L ha<sup>-1</sup> of spray solution at 276 kPa. First, late-POST treatments were applied at V7 growth stage (Palmer amaranth plants were 70-90 cm tall and majority of them were at inflorescence initiation stage) of soybean on July 25, 2022; whereas, sequential late-POST treatments were applied 7 days later on August 2, 2022, when soybean was at R1 growth stage and Palmer amaranth was 90-120 cm in nontreated control plots. GR Palmer amaranth control was assessed visually at 7, 14, and 28 days after last POST (DALPOST) on a scale of 0%-100% (where 0% = no control and 100% = complete control/plant death). Control ratings were based on chlorosis, stunting, and/or necrosis of treated GR Palmer amaranth plants compared to nontreated plants. At 28 DALPOST, Palmer amaranth plants were manually harvested at the soil level using a 1 m<sup>2</sup> quadrat from the center of each plot. The harvested samples from each plot were oven-dried at 65°C for 5 days to determine aboveground shoot dry weights. Palmer amaranth shoot dry weights from herbicide-treated plots were

OPEN ACCESS Agrosystems, Geosciences & Environment AMS 3% + NIS 0.5% AMS 3% + NIS 0.5% AMS 3% + NIS 0.5% (% vol/vol) Adjuvant **AMS 3% AMS 3% AMS 3% AMS 3% AMS 3% AMS 3%** BASF Corp., Syngenta Crop BASF Corp., UPL NA Inc. BASF Corp., FMC Corp. Manufacturer BASF Corp. BASF Corp. BASF Corp. BASF Corp. BASF Corp. BASF Corp. Protection Liberty 280 SL + Dual II Magnum Liberty 280 SL + Anthem Max Liberty 280 SL + Ultra Blazer Liberty 280 SL **Frade name** V7 *fb* R1 V7 fb R1 V7 *fb* R1 Timing (SGS) 77 77 77 ٢٧ 5 5 655 + 128 + 132Rate (g ha<sup>-1</sup>) 655 + 1,337655 *fb* 594 737 fb 594 655 + 280594 fb 594 655 594 737 Glufosinate + pyroxasulfone + Glufosinate + S-metolachlor Glufosinate fb glufosinate Glufosinate fb glufosinate Glufosinate fb glufosinate Glufosinate + acifluorfen Glufosinate Glufosinate Glufosinate Herbicide

List of herbicide treatments, rates, application timing, trade names, manufacturers, and adjuvants used in field study to evaluate the effect of late-season application of glufosinate-based

programs on glyphosate-resistant Palmer amaranth control in 2022 growing season at Kansas State University Agricultural Research Center near Hays, Kansas.

**FABLE 1** 

Abbreviations: AMS, ammonium sulfate; *fb*, followed by; NIS, nonionic surfactant; SGS, soybean growth stage

3 of 8

AMS 3% + NIS 0.5%

Bayer CropScience

Roundup PowerMax

V7 fb R1

1,260 fb 1,260

Glyphosate fb glyphosate

fluthiacet-methyl

expressed as a percent shoot dry weight reduction relative to the nontreated plots using Equation 1:

$$Y = \left[ \left( A - B \right) / A \right] \times 100 \tag{1}$$

where *Y* represents GR Palmer amaranth shoot dry weight reduction (%), *A* is the averaged shoot dry weights from nontreated plots, and *B* is the shoot dry weight from herbicide-treated plot. Soybean grain yields (kg ha<sup>-1</sup>) were estimated by harvesting the middle two rows from each plot using a plot combined at maturity. Soybean grain yields were adjusted to 13% moisture content.

#### 2.2 | Statistical analysis

Data were subjected to analysis of variance (ANOVA) using the PROC MIXED procedure in SAS 9.3. Data were checked for ANOVA assumptions (normality of residuals and homogeneity of variance) using PROC UNIVARIATE in SAS (SAS Institute), and all data met those assumptions. The ANOVA model included all late-POST glufosinate treatments as fixed effects and replications as random effects. Data on GR Palmer amaranth control (%) from nontreated plots were excluded from the analyses. Treatment means were separated using a Fisher's protected LSD test (p < 0.05).

### **3** | RESULTS AND DISCUSSION

The 2022 growing season at the study site had relatively warmer and drier weather conditions than a typical growing season. Mean monthly air temperatures of 24, 27, 26, 22, and 14°C were observed in June, July, August, September, and October, respectively (Figure 1). The average daily relative humidity on July 25 (first late-POST application) and August 2 (sequential late-POST application) was 76% and 45%, respectively (Figure 1). Below-average precipitation occurred in 2022, with total monthly precipitation of 36, 45, 35, 54, and 4 mm in June, July, August, September, and October, respectively (Figure 1). The soil moisture was below normal, particularly during the reproductive growth stages of soybean.

# **3.1** | Palmer amaranth control and shoot dry weights reduction

GR Palmer amaranth control was significantly influenced by late-POST programs at 7, 14, and 28 DALPOST ( $p \le 0.05$  for each). At 7 DALPOST, all glufosinate-based programs provided 84%–95% control (Table 2). With a single late-POST application at V7 soybean growth stage, control improved

from 84% to 92% as glufosinate rate increased from 594 g ha<sup>-1</sup> to 737 g ha<sup>-1</sup>. A sequential late-POST glufosinate application at 594 g ha<sup>-1</sup> during R1 soybean growth stage improved GR Palmer amaranth control by 11% and 6% following a previous glufosinate treatment at 594 g ha<sup>-1</sup> or 655 g ha<sup>-1</sup> applied at V7 soybean growth stage, respectively. Besides, tank-mix application of residual herbicides with 655 g ha<sup>-1</sup> of glufosinate at V7 soybean growth stage provided similar control of GR-Palmer amaranth as two sequential late-POST glufosinate applications. Vann et al. (2017) also observed no glufosinate rate differences for Palmer amaranth control with two sequential glufosinate applications. Similar treatment differences were observed at 14 DALPOST with Palmer amaranth control varied from 83% to 95% with all glufosinate-based treatments. By 28 DAL-POST, Palmer amaranth control was still >90% with two late-POST glufosinate treatments made at V7 followed by R1 soybean stages. This level of control was higher than a single late-POST glufosinate treatment at 594 g ha<sup>-1</sup> or where a glufosinate + S-metolachlor tank mixture was used at the V7 soybean stage (Table 2). With a single V7 stage application, GR Palmer amaranth control was similar regardless of glufosinate rates. Similarly, Vann et al. (2017) reported disappearance of glufosinate rate effect with time for the control of 5- to 35-cm tall Palmer amaranth. As expected, glyphosate had little effect on GR Palmer amaranth with  $\leq 12\%$  control after two sequential treatments at 1260 g ha<sup>-1</sup> at V7 *fb* R1 soybean growth stage.

Glufosinate applied late-POST in this study was highly effective, and provided >82% control of tall (70-90 cm) GR Palmer amaranth. Norsworthy et al. (2008) reported  $\geq$ 99% control of GR-Palmer amaranth with glufosinate applied when Palmer amaranth plants were at six leaves growth stage. In Connecticut, glufosinate applied at 590 g ha<sup>-1</sup> provided complete control of a multiple herbicide-resistant Palmer amaranth (Aulakh et al., 2021). Other researchers found sequential applications of glufosinate were more effective than a single application for controlling broadleaf and grass weeds (Aulakh & Jhala, 2015; Hoffner et al., 2012; Jhala et al., 2017; Wiesbrook et al., 2001). Moreover, glufosinate efficacy on weeds also varies with the application time of the day. Palmer amaranth was controlled 97% when glufosinate was applied mid-day compared with 63% control when glufosinate was applied at sunrise (Copeland et al., 2019). Coetzer et al. (2001) observed relative humidity and air temperature-dependent variation in Palmer amaranth control by glufosinate. They noted that Palmer amaranth control was improved by 17%-35% 1 day after glufosinate application as the relative humidity increased from 35% to 90%.

Shoot dry weight reduction of GR Palmer amaranth was consistent with the control results at 28 DALPOST (Table 1). Late-POST glufosinate-based programs, except for a single application of glufosinate at 594 g ha<sup>-1</sup> at V7 soybean growth



**FIGURE 1** Daily air temperature (C), relative humidity (%), and precipitation (mm) during the 2022 soybean growing season at Kansas State University Agricultural Research Center near Hays, Kansas.

stage, were similar with 83%–91% reduction in GR Palmer amaranth shoot dry weights compared to the nontreated control. Similar reductions in weed biomass with glufosinate have previously been reported (Aulakh & Jhala, 2015; Vann et al., 2017).

#### 3.2 | Soybean grain yield

Late-POST glufosinate-based programs resulted in higher soybean grain yields than the nontreated control or a glyphosate fb glyphosate. The highest soybean yield (701 kg  $ha^{-1}$ ) was observed with 737 g  $ha^{-1}$  of glufosinate at V7 soybean growth stage fb 594 g ha<sup>-1</sup> of glufosinate at R1 soybean growth stage. This level of soybean grain yield was higher than observed with glufosinate + aciflourfen, glufosinate + S-metolachlor, glyphosate fb glyphosate, and the nontreated control with 470, 444, 270, and 218 kg  $ha^{-1}$ respectively. Although the GR Palmer amaranth control was similar to other glufosinate-based programs, the glufosinate + acifluorfen or S-metolachlor tank-mix had an adverse impact on soybean yield. This was perhaps due to higher soybean injury (approximately 15%-18%) from these programs compared to <6% injury with all other glufosinate-based programs (data not shown). Soybean injury from aciflourfen was characterized as necrosis, chlorosis, and height reduction. Aciflourfen was found to be highly injurious (>20%) to soybean (Aulakh et al., 2016). In another study, Aulakh and Jhala (2015) observed approximately 20% injury to soybean with an EPOST application of glufosinate + acetochlor or glufosinate + S-metolachlor tank mixture. However, observed EPOST

injury in their research was transitory and therefore, had no impact on soybean grain yield.

Lower than normal soybean grain yields were produced in this study. Typical grain yield for this soybean maturity group at the KSU-ARC is 1512 kg ha<sup>-1</sup> (Kumar, Liu, Peterson, et al., 2021). Below average soybean grain yields in current study could be attributed to the exceptionally hot and dry weather conditions at reproductive growth stage during 2022. The cumulative precipitation for the 2022 soybean-growing season was 261 mm as compared to the historic average of 406 mm.

## 4 | CONCLUSIONS AND PRACTICAL IMPLICATIONS

GR Palmer amaranth presents a serious challenge to soybean production because effective herbicides in soybean are dwindling. Glufosinate-resistant soybean was first released for large-scale commercial cultivation in 2009, although limited cultivation had already begun in 1999 (Wiesbrook et al., 2001). Glufosinate is a POST, contact herbicide for control of emerged broadleaf, and grass weeds in glufosinateresistant soybean (Aulakh & Jhala, 2015; Haas & Muller, 1987; Jhala et al., 2017). In this study, several late-POST glufosinate-based herbicide programs effectively controlled ( $\geq$ 82%) 70–90 cm tall GR Palmer amaranth. Single late-POST glufosinate application at  $\geq$ 655 g ha<sup>-1</sup> during the V7 soybean growth stage was similar to two late-POST applications at V7 and R1 soybean growth stages. Consequentially, glufosinate-based programs produced similar soybean yields,

| Herbicide                                      | Rate (g ha $^{-1}$ ) | 7 DALPOST | 14 DALPOST    | 28 DALPOST | Shoot dry weight reduction | Grain yield (kg<br>ha <sup>-1</sup> ) |
|--|----------------------|-----------|---------------|------------|----------------------------|---------------------------------------|
|  |                      |           | — % control — |            | % of no                    | ontreated                             |
| Nontreated                                     | -                    | -         | -             | -          | -                          | 218 d                                 |
| Glufosinate                                    | 594                  | 84 e      | 83 d          | 82 c       | 75 b                       | 562 ab                                |
| Glufosinate                                    | 655                  | 88 cd     | 88 c          | 87 abc     | 83 a                       | 539 ab                                |
| Glufosinate                                    | 737                  | 92 ab     | 91 abc        | 88 abc     | 85 a                       | 558 ab                                |
| Glufosinate <i>fb</i> glufosinate              | 594 <i>fb</i> 594    | 95 a      | 95 a          | 91 ab      | 91 a                       | 626 a                                 |
| Glufosinate <i>fb</i> glufosinate              | 655 <i>fb</i> 594    | 94 a      | 93 abc        | 92 ab      | 89 a                       | 650 a                                 |
| Glufosinate <i>fb</i> glufosinate              | 737 fb 594           | 95 a      | 94 ab         | 93 a       | 89 a                       | 701 a                                 |
| Glufosinate +<br>acifluorfen                   | 655 + 280            | 90 bc     | 89 bc         | 89 abc     | 86 a                       | 470 bc                                |
| Glufosinate +<br>S-metolachlor                 | 655 + 1,337          | 92 ab     | 88 c          | 84 bc      | 84 a                       | 444 bc                                |
| Glufosinate +<br>pyroxasulfone +<br>fluthiacet | 655 + 128 + 132      | 93 ab     | 92 abc        | 89 abc     | 86 a                       | 524 ab                                |
| Glyphosate <i>fb</i> glyphosate                | 1,260 fb 1,260       | 10 e      | 12 e          | 11 d       | 33 c                       | 270 d                                 |

**TABLE 2** Effect of late-season applied glufosinate-based programs on glyphosate-resistant Palmer amaranth control, shoot dry weight reduction and grain yields of glyphosate/dicamba/glufosinate-resistant soybean in no-till dryland.

*Note*: Means within each column followed by similar alphabets are not different according to Fisher's Protected LSD (p < 0.05) test.

Abbreviations: DALPOST, days after last POST; *fb*, followed by, POST, postemergence.

which were higher, compared to the nontreated control or glyphosate fb glyphosate. Mixing glufosinate with pyroxasulfone + fluthiacet at V7 soybean growth stage provided similar soybean grain yield compared with single or sequential late-POST glufosinate applications. Contrary to previous findings, a late-POST aciflourfen or *S*-metolachlor mixed with glufosinate caused severe injury to soybean that resulted in reduced soybean yields than a glufosinate + pyroxasulfone + fluthiacet.

Recently, Palmer amaranth populations resistant to ALSinhibitors, glufosinate, glyphosate, HPPD-inhibitor, PPOinhibitor, and synthetic auxins herbicides have been documented in multiple states in the United States (Aulakh et al., 2021; Chahal et al., 2015, 2018; Foster & Steckel, 2022; Heap, 2023; Kumar et al., 2019; Priess et al., 2022; Salas et al., 2016; Varanasi et al., 2018). Evolution of multiple herbicide resistance in Palmer amaranth would defy chemical control even in crops with stacked gene herbicide resistant trait technologies. Therefore, future research efforts should investigate combination of chemical control with ecological weed management tactics such as competitive crop rotation, cover crop, planting densities, tillage, and harvest weed seed control (HWSC) techniques (weed seed destructor and chaff lining) for managing GR Palmer amaranth. In this context, a multi-state (Iowa, Kansas, and Arkansas) field research is underway to investigate the long-term impact of multitactic approaches (cover crops, herbicides, and HWSC techniques) on GR pigweeds seedbanks. Future research should also investigate the weed control efficacy and soybean safety of late-POST glufosinate + PPO-inhibitor herbicides tank-mixes under different relative humidity and temperature regimes. Tank mixing residual herbicides with a viable late-season POST treatment is often recommended for safeguarding herbicide-resistant crop technologies and managing herbicide-resistant weeds (Norsworthy et al., 2012). Addition of a late-POST residual herbicide offer added advantages, for example, diversity of herbicide sites of action which will alleviate the selection pressure of a single herbicide (Diggle et al., 2003; Johnson et al., 2012) and reduce the weed seed bank in the soil (Legleiter et al., 2009).

#### AUTHOR CONTRIBUTIONS

Vipan Kumar: Conceptualization, data curation, formal analysis, investigation, methodology, project administration, resources, software, supervision, validation, visualization, writing–original draft. Jatinder Aulakh: Data curation, validation, writing–original draft, writing–review and editing. Rui Liu: Investigation, methodology, validation, writing– review and editing. Amit Jhala: Conceptualization, validation, visualization, writing–review and editing.

#### ACKNOWLEDGMENTS

The authors would like to thank Mr. Taylor Lambert, Mr. Sachin Dhanda, and Mr. Matt Vredenburg at Kansas State University Agricultural Research Center, Hays, KS, for providing technical support in conducting this research. This research received no specific grant from any funding agency or the commercial or not-for-profit sectors.

### CONFLICT OF INTEREST STATEMENT

The authors declare no conflicts of interest.

## ORCID

Vipan Kumar (1) https://orcid.org/0000-0002-8301-5878

## REFERENCES

- Adhikary, D., & Pratt, D. B. (2015). Morphologic and taxonomic analysis of the weedy and cultivated *Amaranthus hybridus* species complex. *Systematic Botany*, 40, 604–610. https://doi.org/10.1600/ 036364415X688376
- Anderson, D. M., Swanton, C. J., Hall, J. C., & Mersey, B. G. (1993). The influence of temperature and relative humidity on the efficacy of glufosinate-ammonium. *Weed Research*, 33(2), 139–147. https://doi. org/10.1111/j.1365-3180.1993.tb01927.x
- Anonymous (2019). Liberty 280 SL herbicide product label. EPA Reg. No. 7969-448. (p. 21) Research Triangle Park, NC: BASF Corporation.
- Aulakh, J. S., Chahal, P. S., & Jhala, A. J. (2016). Glyphosateresistant weed control and soybean injury in response to different PPO-inhibiting herbicides. *Journal of Agricultural Sciences*, 8, 1–8.
- Aulakh, J. S., Chahal, P. S., Kumar, V., Price, A. J., & Guillard, K. (2021). Multiple herbicide-resistant Palmer amaranth (*Amaranthus palmeri*) in Connecticut: Confirmation and response to POST herbicides. Weed Technology, 35, 457–463. https://doi.org/10.1017/wet.2021.6
- Aulakh, J. S., & Jhala, A. J. (2015). Comparison of glufosinate-based herbicide programs for broad-spectrum weed control in glufosinateresistant soybean. *Weed Technology*, 29, 419–430. https://doi.org/10. 1614/WT-D-15-00014.1
- Brabham, C., Norsworthy, J. K., Houston, M. M., Varanasi, V. K., & Barber, T. (2019). Confirmation of S-metolachlor resistance in Palmer amaranth (*Amaranthus palmeri*). Weed Technology, 33(5), 720–726. https://doi.org/10.1017/wet.2019.44
- Chahal, P. S., Aulakh, J. S., Jugulam, M., & Jhala, A. J. (2015). Herbicide-resistant Palmer amaranth (*Amaranthus palmeri* S. Wats.) in the United States: Mechanisms of resistance, impact, and management. In A. Price, J. Kelton, & L. Sarunaite (Eds.), *Herbicides, agronomic crops and weed biology* (pp. 1–29). In Tech.
- Chahal, P. S., Irmak, S., Gaines, T., Amundsen, K., Jugulam, M., Jha, P., Travlos, I. S., & Jhala, A. J. (2018). Control of photosystem II–and 4-hydroxyphenylpyruvate dioxygenase inhibitor–resistant Palmer amaranth (*Amaranthus palmeri*) in conventional corn. Weed Technology, 32, 326–335. https://doi.org/10.1017/wet.2017.111
- Coetzer, E., Al-Khatib, K., & Loughin, T. M. (2001). Glufosinate efficacy, absorption, and translocation in amaranth as affected by relative humidity and temperature. *Weed Science*, 49, 8–13. https://doi.org/10. 1614/0043-1745(2001)049%5b0008:GEAATI%5d2.0.CO;2
- de Sanctis, J. H. S., Knezevic, S. Z., Kumar, V., & Jhala, A. J. (2021). Effect of single or sequential POST herbicide applications on seed

production and viability of glyphosate-resistant Palmer amaranth (*Amaranthus palmeri*) in dicamba- and glyphosate-resistant soybean. *Weed Technology*, *35*(3), 449–456. https://doi.org/10.1017/wet.2021. 7

- Diggle, A. J., Neve, P. B., & Smith, F. P. (2003). Herbicides used in combination can reduce the probability of herbicide resistance in finite weed populations. *Weed Research*, 43, 371–382. https://doi.org/10. 1046/j.1365-3180.2003.00355.x
- Drake Copeland, J., Montgomery, G. B., & Steckel, L. E. (2019). Evaluation of the time-of-day effect of herbicides applied POST on protoporphyrinogen IX oxidase-resistant and -susceptible Palmer amaranth (*Amaranthus palmeri*). Weed Technology, 33, 651–657. https://doi.org/10.1017/wet.2019.43
- Foster, D. C., & Steckel, L. E. (2022). Confirmation of dicamba-resistant Palmer amaranth in Tennessee. *Weed Technology*, *36*(6), 777–780. https://doi.org/10.1017/wet.2022.87
- Haas, P., & Muller, F. (1987). Behaviour of glufosinate-ammonium in weeds. *Proceedings of the 10th BCPC Congress—Weeds*, UK, 1075– 1082.
- Heap, I. M. (2023). International survey of herbicide resistant weeds. http://www.weedscience.org
- Hinchee, M. A. W., Padgette, S. R., Kishore, G. M., Delannay, X., & Fraley, R. T. (1993). Herbicide-tolerant crops. In S. Kung, & R. Wu (Eds.), *Transgenic Plants* (pp. 243–263). Academic Press.
- Hoffner, A. E., Jordan, D. L., Chandi, A., York, A. C., Dunphy, E. J., & Everman, W. J. (2012). Management of Palmer amaranth (*Amaranthus palmeri*) in glufosinate resistant soybean (*Glycine max*) with sequential applications of herbicides. *International Scholarly Research Notices*, 2012, 1–7.
- Horak, M. J., & Loughin, T. M. (2000). Growth analysis of four Amaranthus species. Weed Science, 48, 347–355. https://doi.org/10.1614/ 0043-1745(2000)048%5b0347:GAOFAS%5d2.0.CO;2
- Jenkins, J. (2023). EPA adjusts dicamba cutoff dates in '23. DTN/Progressive Farmer. https://www.dtnpf.com/agriculture/web/ ag/crops/article/2023/02/16/dicamba-labels-now-list-june-12-ia
- Jha, P., & Norsworthy, J. K. (2009). Soybean canopy and tillage effects on emergence of Palmer amaranth (*Amaranthus palmeri*) from a natural seed bank. *Weed Science*, 57, 644–651. https://doi.org/10.1614/ WS-09-074.1
- Jhala, A. J., Sandell, L. D., Sarangi, D., Kruger, G. R., & Knezevic, S. Z. (2017). Control of glyphosate-resistant common waterhemp (*Amaranthus rudis*) in glufosinate-tolerant soybean. *Weed Technology*, 31, 32–45. https://doi.org/10.1017/wet.2016.8
- Johnson, G., Breitenbach, F., Behnken, L., Miller, R., Hoverstad, T., & Gunsolus, J. (2012). Comparison of herbicide tactics to minimize species shifts and selection pressure in glyphosate-resistant soybean. *Weed Technology*, 26, 189–194. https://doi.org/10.1614/WT-D-11-00106.1
- Jones, E., Leon, R. G., & Everman, W. J. (2022). Biological effects on Palmer amaranth surviving glufosinate. *Agrosystems, Geosciences & Environment*, 5(4), e20315. htts://doi.org/10.1002/agg2.20315
- Keeley, P. E., Carter, C. H., & Thullen, R. J. (1987). Influence of planting date on growth of Palmer amaranth (*Amaranthus palmeri*). Weed Science, 1, 199–204. https://doi.org/10.1017/S0043174500079054
- Klingaman, T. E., & Oliver, L. R. (1994). Palmer amaranth (*Amaranthus palmeri*) interference in soybeans (*Glycine max*). Weed Science, 42, 523–527. https://doi.org/10.1017/S0043174500076888
- Kumar, V., Liu, R., Boyer, G., & Stahlman, P. W. (2019). Confirmation of 2,4-D resistance and identification of multiple resistance

in a Kansas Palmer amaranth (*Amaranthus palmeri*) population. Pest Management Science, 75, 2925–2933. https://doi.org/10.1002/ ps.5400

- Kumar, V., Liu, R., Jhala, A. J., Jha, P., & Manuchehri, M. (2021). Palmer amaranth (*Amaranthus palmeri*) control in postharvest wheat stubble in the Central Great Plains. *Weed Technology*, 35(6), 945–949. https:// doi.org/10.1017/wet.2021.64
- Kumar, V., Liu, R., Peterson, D. E., & Stahlman, P. W. (2021). Effective two-pass herbicide programs to control glyphosate-resistant Palmer amaranth (*Amaranthus palmeri*) in glyphosate/dicamba-resistant soybean. Weed Technology, 35, 128–135. https://doi.org/10.1017/wet. 2020.90
- Kumar, V., Liu, R., & Stahlman, P. W. (2020). Differential sensitivity of Kansas Palmer amaranth populations to multiple herbicides. *Agronomy Journal*, 112, 2152–2163. https://doi.org/10.1002/agj2. 20178
- Legleiter, T. R., Bradley, K. W., & Massey, R. E. (2009). Glyphosate resistant waterhemp control and economic returns with herbicide treatments in soybean. *Weed Technology*, 23, 54–61. https://doi.org/ 10.1614/WT-08-069.1
- Liu, R., Kumar, V., Jha, P., & Stahlman, P. W. (2022). Emergence pattern and periodicity of Palmer amaranth (*Amaranthus palmeri*) populations from southcentral Great Plains. *Weed Technology*, 36(1), 110–117. https://doi.org/10.1017/wet.2021.81
- Meyer, C. J., Norsworthy, J. K., Young, B. G., Steckel, L. E., Bradley, K. W., Johnson, W. G., Loux, M. M., Davis, V. M., Kruger, G. R., Bararpour, M. T., Ikley, J. T., Spaunhorst, D. J., & Butts, T. R. (2015). Herbicide program approaches for managing glyphosate-resistant Palmer amaranth (*Amaranthus palmeri*) and waterhemp (*Amaranthus tuberculatus* and *Amaranthus rudis*) in future soybean-trait technologies. *Weed Technology*, 29, 716–729. https://doi.org/10.1614/WT-D-15-00045.1
- Norsworthy, J. K., Griffith, G. M., Scott, R. C., Smith, K. L., & Oliver, L. R. (2008). Confirmation and control of glyphosate-resistant Palmer amaranth (*Amaranthus palmeri*) in Arkansas. *Weed Technology*, 22, 108–113. https://doi.org/10.1614/WT-07-128.1
- Norsworthy, J. K., Ward, S. M., Shaw, D. R., Llewellyn, R. S., Nichols, R. L., Webster, T. M., Bradley, K. W., Frisvold, G., Powles, S. B., Burgos, N. R., Witt, W. W., & Barrett, M. (2012). Reducing the risks of herbicide resistance: Best management practices and recommendations. *Weed Science*, 60, 31–62. https://doi.org/10.1614/WS-D-11-00155.1
- Priess, G. L., Norsworthy, J. K., Godara, N., Mauromoustakos, A., Butts, T. R., Roberts, T. L., & Barber, T. (2022). Confirmation of glufosinate-resistant Palmer amaranth and response to other herbicides. *Weed Technology*, *36*, 368–372. https://doi.org/10.1017/wet. 2022.21

- Salas, R. A., Burgos, N. R., Tranel, P. J., Singh, S., Glasgow, L., Scott, R. C., & Nichols, R. L. (2016). Resistance to PPO-inhibiting herbicide in Palmer amaranth from Arkansas. *Pest Management Science*, 72, 864–869. https://doi.org/10.1002/ps.4241
- Sauer, J. (1957). Recent migration and evolution of the dioecious amaranths. Evolution; Internation Journal of Organic Evolution, 11, 11–31. https://doi.org/10.2307/2405808
- Steckel, L. E., Sprague, C. L., Stoller, E. W., & Wax, L. M. (2004). Temperature effects on germination of nine *Amaranthus* species. *Weed Science*, 52, 217–221. https://doi.org/10.1614/WS-03-012R
- Vann, R. A., York, A. C., Cahoon, C. W., Buck, T. B., Askew, M. C., & Seagroves, R. W. (2017). Glufosinate plus dicamba for rescue Palmer amaranth control in XtendFlex cotton. *Weed Technology*, *31*, 666– 674. https://doi.org/10.1017/wet.2017.68
- Van Wychen, L. (2017). Survey of the most common and troublesome weeds in grass crops, pasture and turf in the United States and Canada. Weed Science Society of America National Weed Survey Dataset. http://wssa.net/wp-content/uploads/WSSA-Weed-Survey-2017-05-19.pdf
- Varanasi, V. K., Brabham, C., Norsworthy, J. K., Nie, H., Young, B. G., Houston, M., Barber, T., & Scott, R. C. (2018). A statewide survey of PPO-inhibitor resistance and the prevalent target-site mechanisms in Palmer amaranth (*Amaranthus palmeri*) accessions from Arkansas. *Weed Science*, 66, 149–158. https://doi.org/10.1017/wsc.2017.68
- Ward, S. M., Webster, T. M., & Steckel, L. E. (2013). Palmer amaranth (*Amaranthus palmeri*): A review. Weed Technology, 27, 12–27. https://doi.org/10.1614/WT-D-12-00113.1
- Wiesbrook, M. L., Johnson, W. G., Hart, S. E., Bradley, P. R., & Wax, L. M. (2001). Comparison of weed management systems in narrow-row, glyphosate and glufosinate-resistant soybean (*Glycine max* L.). Weed *Technology*, 15, 122–128. https://doi.org/10.1614/0890-037X(2001) 015%5b0122:COWMSI%5d2.0.CO;2

**How to cite this article:** Kumar, V., Aulakh, J. S., Liu, R., & Jhala, A. J. (2023). Late postemergence glufosinate-based programs for glyphosate-resistant Palmer amaranth control in

dicamba/glufosinate/glyphosate-resistant soybean. Agrosystems, Geosciences & Environment, 6, e20378. https://doi.org/10.1002/agg2.20378